Two new species of *Panaeolus* (Psathyrellaceae, Agaricales) from coprophilous habitats of Punjab, India

Amandeep Kaur^{1*}, NS Atri² and Munruchi Kaur²

¹Desh Bhagat College of Education, Bardwal–Dhuri–148024, Punjab, India. ²Department of Botany, Punjabi University, Patiala–147002, Punjab, India.

(Received on: 06 June, 2014; accepted on: 02 July, 2014)

ABSTRACT

Two new coprophilous species of *Panaeolus*, namely *P. cyanoannulatus* and *P. lepus-stercus*, of family *Psathyrellaceae* are described from Punjab, India. *Panaeolus cyanoannulatus* was collected on a mixed cow and horse dung heap from Hoshiarpur district and *P. lepus-stercus* was located growing scattered on rabbit pellets from Pathankot district of Punjab state in India. *Panaeolus cyanoannulatus* is a blue staining mushroom with annulate stipe while *P. lepus-stercus* is characterized by yellowish gray umbonate pileus, 2– and 4–spored basidia, limoniform hexagonal basidiospores, polymorphic chrysocystidia and absence of pileocystidia and clamp connections. In this paper, these two taxa are described, illustrated, and compared with similar species.

Key Words: Agaricomycetes, Basidiomycota, blue staining, mushrooms, taxonomy.

INTRODUCTION

The genus *Panaeolus* (Fr.) Quél. is represented by 15 species the world over (Kirk *et al.* 2008). However, MycoBank mentions 150 legitimate records of *Panaeolus*. The genus is characterized by carpophores which are often bluing when bruised or with age; adnexed to adnate, variegated, grayishblack lamellae; epithelial pileus cuticle and reddish brown to blackish brown spores which do not fade in concentrated sulphuric acid.

From India 24 taxa are already known (Bose 1920, Pathak & Ghosh 1962, Ghosh *et al.* 1967, Sathe & Sasangan 1977, Sarbhoy & Daniel 1981, Natarajan & Raaman 1983, 1984, Bhide *et al.* 1987, Abraham 1991, Dhancholia *et al.* 1991, Lakhanpal 1993, 1995, Bhavani Devi 1995, Patil *et al.* 1995, Vrinda *et al.* 1999, Manimohan *et al.* 2007, Amandeep *et al.* 2013). Based on the survey conducted to various dung localities of Punjab, collections assigned to *P. cyanoannulatus* and *P. lepus–stercus* were collected and described as new species.

Corresponding author: amandeepbotany75@gmail.com

MATERIAL AND METHODS

The materials were collected from dung localities in Punjab. The macroscopic characters pertaining to gross morphology, shape, color and size of pileus, stipe, etc. were noted down from the fresh material on the field key especially designed for the purpose (Atri et al. 2005). The color terminology used is that of Kornerup & Wanscher (1978). The specimens were hot air dried and packed in cellophane paper 1–4 bags containing dichlorobenzene. The microscopic structures were observed by cutting free hand sections after reviving a part of the dried materials in 10% KOH solution and staining the sections in 0.16% Cotton blue. Line drawings of microscopic details were drawn with the aid of Camera lucida under oil immersion lens. The collections have been deposited in the Herbarium of Botany Department, Punjabi University, Patiala (Punjab), India under PUN (Holmgren & Keuken 1974). The photographs and microscopic details are given in Figs. 1-2 for P. cyanoannulatus and Figs. 3–4 for *P. lepus–stercus*.

TAXONOMIC DESCRIPTIONS

Panaeolus cyanoannulatus Atri, M. Kaur & A.Kaur, sp. nov.gs. 1-2

MycoBank no.: MB 805219.

Etymology: The name of the species is based on the development of blue stains on bruising and annulate stipe character.

Diagnosis: It is a typical blue staining species with prominently annulate stipe. It differs from the other *Panaeolus* species in possessing annulate stipe and lacking chrysocystidia within the hymenium.

Carpophores 6.5–7.3 cm in height. Pileus 1.3–1.4 cm broad, 1.3–1.5 cm high, conical with pointed umbo; surface yellowish brown $(5E_8)$, staining bluish when bruised, moist, smooth; margin regular, not splitting at maturity, non–striate; cuticle fully–peeling; flesh thin, becoming bluish on exposure; taste and odor mild. Lamellae broadly adnate, unequal, 3–sized, sub–distant, narrow, 0.2–0.25 cm broad, fragile, bluish yellow to bluish brown, finally bluish black; gill edges smooth; spore print black. Stipe 6.4–7.2 cm long, 0.1 cm broad, tubular, equal in diameter throughout, hollow, surface yellowish brown (5E₈), bluing when handled, smooth; annulate, annulus single, ring like, membranous, attached near the centre.

Basidiospores 13–15.6 \times 7–10 μ m (Q = 1.68), elongated ellipsoidal in face view, slightly flattened in side view, with a broad central germ pore, thick-walled, smooth, dark brown, not bleaching in concentrated H₂SO₄. Basidia 14–18.5 \times 7-9 µm, cylindrical to clavate, 2-, 4-spored, thin walled, hyaline; sterigmata 2.8-5.7 µm long. Gill edges sterile. Cheilocystidia $14-23 \times 2.7-4.3 \mu m$, abundant, cylindrical to clavate, with inflated apex, thin-walled, hyaline, apical region 4.3-7 µm broad. Pleurocystidia absent. Pileus cuticle a four to five layered stratified cellular epithelium with scattered pileocystidia; cellular elements 8.5-17 µm broad, subglobose to globose, thin-walled, hyaline; pileocystidia $34-71 \times 5-7.5 \mu m$, polymorphic, cylindrical, lageniform, narrow, wavy, some with subcapitate apex, thin-walled, hyaline; context hyphae interwoven, thin walled, hyaline $6.7-18.7 \,\mu m$ broad. Hymenophoral trama regular composed of thin-walled, 5.7-14 µm broad hyphae. Subhymenium pseudoparenchymatous. Stipe cuticle hyphal;

caulocystidia present, similar to cheilocystidia in shape but larger in size measuring $24-45 \times 4.4-6.4$ µm, arranged in scattered tufts, thin-walled, hyaline, apical region 8–11.4 µm broad; context made up of longitudinally arranged, cylindrical, thin-walled 5.7– 17 µm broad hyphae. Clamp connections present in the stipe context hyphae.

Material examined: India, Punjab, Hoshiarpur, Jeewanpur Jattan (295 m), growing in group on mixed cow and horse dung heap in pasture land, 18 July 2008, Amandeep Kaur, PUN 4223 (Holotype).

Remarks: The above examined collection is a bluestaining Panaeolus species with well developed annulus on the stipe. It can be distinguished from the other blue-staining allied Panaeolus species including P. cyanescens (Berk. & Br.) Sacc., P. tropicalis Oláh, P. cambodginiensis Oláh & R. Heim and P. subbalteatus (Berk. & Br.) Sacc. in being annulate. From P. cvanescens, it also differs in lacking metulloidal cystidia within the hymenium. In P. tropicalis, the spores are 10-12 µm long (Arora 1986) as compared to 13–15.6 µm long in the present collection. Other allied species P. cambodginiensis possesses ochre golden brown to pallid straw colored pileus which is often cracked or wrinkled with comparatively smaller spores measuring $10-12.5 \times$ 6.5-9 µm (Stamets 1996). As compared, the presently examined collection has yellowish brown smooth pileus and larger basidiospores. Although P. subbalteatus is quite close in spore size $(10-14 \times 7-9)$ µm) to the present collection but differs in having exannulate stipe, besides its pileus develops a dark marginal band when it begins to lose moisture and also it occasionally develops a faint blue stain at the base of the stipe (Arora 1986). As compared in the above examined collection the stipe is annulate, the pileus lacks any marginal band and the whole fructification develops bluish tinge when handled. Another species near to the above examined collection is P. semiovatus (Fr.) Lundell & Nannf. which also has an annulus, but it has pale cream to pale buff campanulate cap, chrysocystidia within the hymenium and the spores measuring $16-20 \times 9-11$ µm in size (Watling and Gregory 1987), in comparison to yellowish brown conical umbonate pileus, no chrysocystidia and smaller spores in the presently examined collection. Giving due significance to this blue-staining annulate Panaeolus, a new species P. cyanoannulatus has been described.

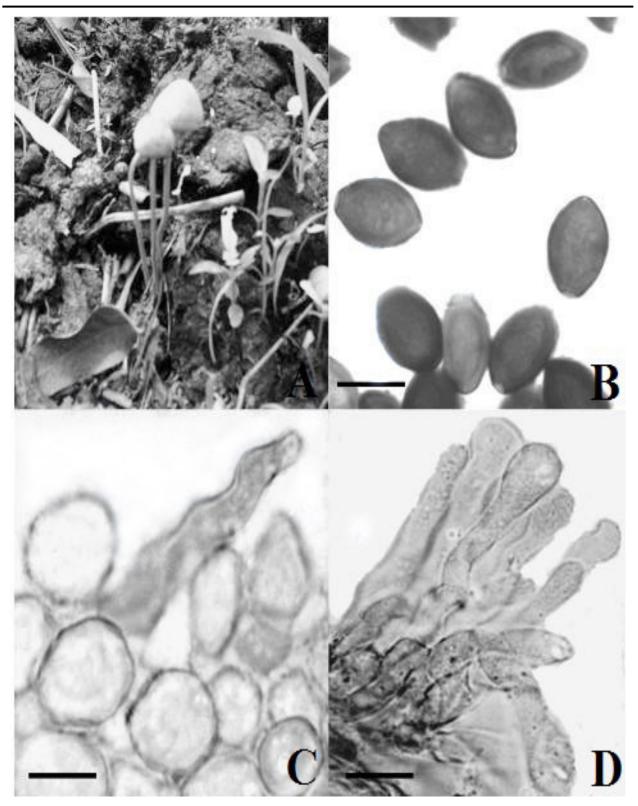


Fig. 1. *Panaeolus cyanoannulatus*. **A-** Carpophores growing in natural habitat; **B-** Basidiospores; **C-** Pileus cuticle elements; **D-** Caulocystidia. *Scale Bars:* B–D 10 μm.

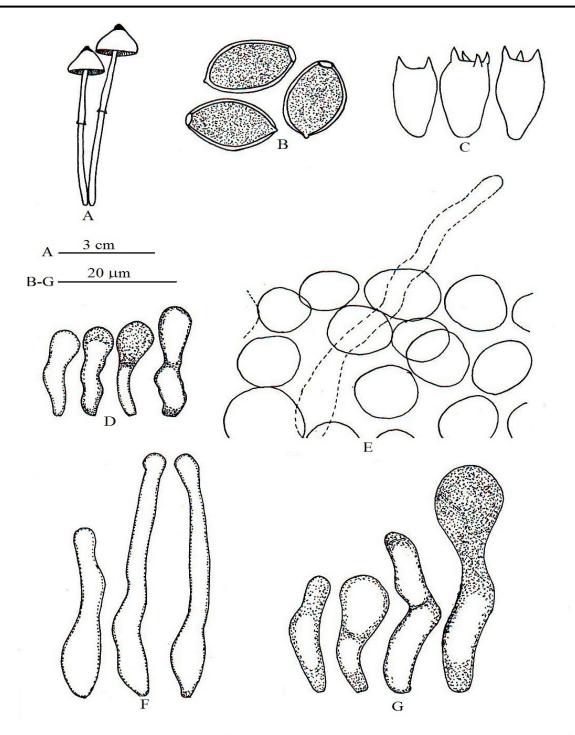


Fig. 2. *Panaeolus cyanoannulatus*. A- Carpophores; B- Basidiospores; C- Basidia; D-Cheilocystidia; E- Pileus cuticle elements; F- Pileocystidia; G- Caulocystidia.

Panaeolus lepus-stercus Atri, M. Kaur & A. Kaur, sp. nov. Figs. 3-4

MycoBank no.: MB 805220.

Etymology: The name of the specific epithet has been drawn from the rabbit on whose pellets the fungus was growing.

Diagnosis: Yellowish gray bluing umbonate pileus, 2– and 4–spored (mostly 2–spored) basidia, large sized limoniform–hexagonal basidiospores, polymorphic chrysocystidia, growing habit on rabbit pellets and absence of pileocystidia and clamp connections are principal diagnostic features.

Carpophores 4.4–4.7 cm in height. Pileus 1– 1.2 cm broad, 0.8–0.9 cm high, campanulate, umbonate; umbo short, pointed; surface yellowish gray (2B₂), bluing when handled, dry, cracked; margin regular, not splitting at maturity, non–striate; cuticle not peeling; flesh thin, yellowish white, unchanging; taste mild, odor not distinctive. Lamellae adnate, unequal, 3–sized, sub–distant, narrow, 0.15–0.2 cm broad, fragile, grayish black; gill edges smooth. Spore print black. Stipe central, 4.3–4.6 cm long, 0.1–0.2 cm broad, tubular, obclavate, solid, surface yellowish gray (2B₂), pruinose; annulus absent.

Basidiospores $13.6-17 \times 10-12 \ \mu m \ (Q =$ 1.4), limoniform to hexagonal in face view, ellipsoidal in side view, with a broad central germ pore, thick-walled, smooth, reddish brown, not bleaching in concentrated H₂SO₄. Basidia 23.8-35.8 \times 12–13.6 µm, clavate, 2–, 4–spored, mostly 2– spored, thin-walled, granular; sterigmata 3.4-6 µm long. Gill edges sterile. Cheilocystidia 23.8-35.8 × 6.8-12 µm, polymorphic, cylindrical, clavate or lageniform, thin-walled, granular, some with densely granular tips. Pleurocystidia chrysocystidioid, 25.5- $53 \times 13.5 - 20.5 \,\mu\text{m}$, polymorphic, ellipsoidal, clavate to ventricose fusoid, thick walled, granular, yellowish brown, some with apical incrustations. Pileus cuticle cellular, cells $17-36 \times 15-26 \mu m$, ovoid, subglobose to clavate, thin-walled, hyaline; pileocystidia absent; pileus context hyphae thin walled, hyaline 8.5-15.3 um broad. Hymenophoral trama regular composed of regular, parallel running, thin-walled, 6.8-17 µm broad hvaline hyphae. Subhymenium pseudoparenchymatous. Stipe cuticle hyphal with scattered caulocystidia; context composed of longitudinally arranged, thin-walled, 8.5-12.7 µm broad hyaline hyphae; caulocystidia $22-43 \times 6-15.3$ um, cylindrical or even lageniform, thin-walled, hyaline to granular. Clamp connections absent throughout.

Material examined: India, Punjab, Pathankot, Sheep and Rabbit Breeding Farm, Dalla Dhar (309 m), growing scattered on rabbit pellets, 01 September 2011, Munruchi Kaur and Amandeep Kaur, PUN 4340 (Holotype).

Remarks: The above examined collection is characterized by yellowish gray umbonate pileus, bisporic as well as tetrasporic basidia, large limoniform–hexagonal spores, polymorphic chrysocystidia, growth on rabbit pellets and absence of pileocystidia and clamp connections.

The species of Panaeolus having chrysocystidia, namely P. tropicalis Oláh, P. ater (J.E. Lange) Kühner & Romagn., P. rubricaulis Petch, P. antillarum (Fr.) Dennis, P. cyanescens (Berk. & Br.) Sacc., P. solidipes (Peck) Sacc. and P. tirunelveliensis Natarajan & Raaman are quite comparable with the presently examined collection. The spores of this collection $(13.6-17 \times 10-12 \text{ }\mu\text{m})$ are much bigger in size in comparison to the spores $(10-12 \times 7-9 \mu m)$ of *P. tropicalis* (Stamets 1996). *P.* ater also mainly differs from it in having smaller spores (10–14 \times 6–8 μ m) with mostly oblique germ pore (Watling and Gregory 1987). Another species P. rubricaulis possesses dark brown pileus with white marginal band and appendiculate margin (Pegler 1986). P. antillarum can be differentiated from the presently examined collection in having much larger spores (16–20 \times 9–12 μ m), areolate pileus surface, stipe bruising brown and chrysocystidia with an irregular amorphous refractive body (Watling and Gregory 1987). Another species near to it is P. cyanescens which differs in having larger carpophores (5–10 cm long) in comparison to smaller carpophores in this species (4.4-4.7 cm long), exumbonate pileus, presence of pileocystidia and clamp connections, and the absence of caulocystidia (Pegler 1986; Wartchow et al. 2010). P. solidipes is different in having large sized pure white carpophores, plano-convex pileus, longitudinally twisted stipe and the presence of clamp connections (Arora 1986). Although P. tirunelveliensis is quite close to the above examined collection in the absence of pileocystidia and clamp connections, but it has only 2-spored basidia, bluish gray exumbonate pileus and the terrestrial habitat. Also the spores (12.6–14 \times 8.4–11.2 μ m) and basidia (16.8–22.4 × 7–8.4 μ m) of P. tirunelveliensis are much smaller in comparison to the spores $(13.6-17 \times 10-12 \ \mu\text{m})$ and basidia (23.8-10) $35.8 \times 12-13.6 \ \mu\text{m}$) in the presently examined collection.

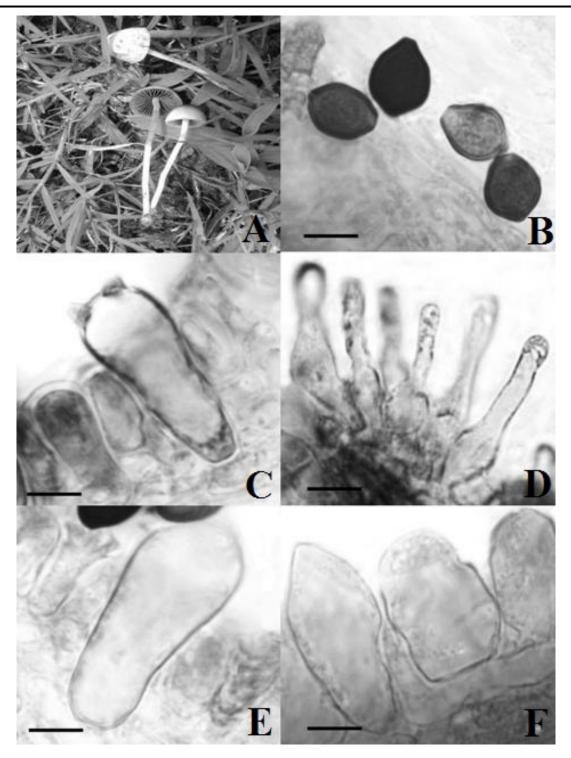


Fig. 3. *Panaeolus lepus–stercus*. **A**- Carpophores growing in natural habitat; **B**- Basidiospores; **C**- A basidium; **D**-Cheilocystidia; **E**- Chrysocystidium; **F**- Pileal elements. *Scale Bars:* B–F 10 μm.

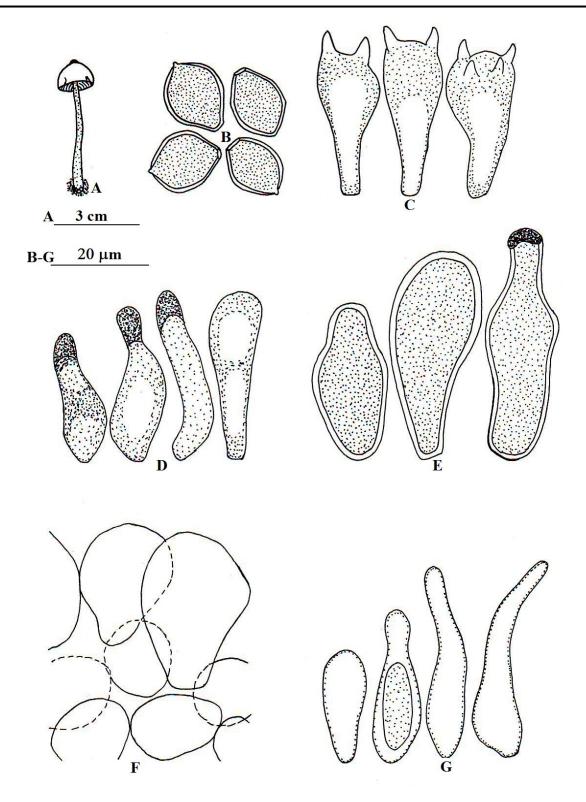


Fig. 4. *Panaeolus lepus–stercus*. A- Carpophore; B- Basidiospores; C- Basidia; D- Cheilocystidia; E- Chrysocystidia; F- Pileus cuticle elements; G- Caulocystidia.

ACKNOWLEDGEMENTS

Authors wish to thank Department of Botany, Punjabi University, Patiala, Punjab for providing laboratory facilities and University Grants Commission for financial assistance under SAP–III programme of which Mycology and Plant Pathology is one of the thrust areas.

REFERENCES

- Abraham SP. 1991. Kashmir fungal flora– an overview. In: Nair MC (ed) Indian Mushrooms, Kerala Agricultural University, Velenikkara, pp 13-24.
- Amandeep K, Atri NS, Munruchi K. 2013. Two new coprophilous varieties of *Panaeolus* (Psathyrellaceae, Agaricales) from Punjab, India. Mycosphere 4: 616-625; doi: 10.5943/mycosphere-4-3-13.
- Arora D. 1986. Mushrooms demystified. A comprehensive guide to the fleshy fungi. Ten Speed Press, Berkeley, CA.
- Atri NS, Kaur A, Kour H. 2005. Wild mushrooms– collection and identification. In: Rai RD, Upadhyay RC, Sharma SR (eds) Frontiers in Mushroom Biotechnology, NRCM, Chambaghat, Solan, Himachal Pradesh, India, pp 9-26.
- Bhavani Devi S. 1995. Mushroom flora of Kerala. In: Chadha KL, Sharma SR (eds) Advances in Hortriculture Vol. 13– Mushrooms, Malhotra Publishing House, New Delhi, pp 277-316.
- Bhide VP, Pande A, Sathe AV, Rad VG, Patwardan, PG. 1987. Fungi of Maharashtra. Maharashtra Association for the Cultivation of Science. MACS Research Institute, Pune, India.
- Bose SR. 1920. Records of Agaricaceae from Bengal. The J Asiatic Soc Bengal 16: 347-354.
- Dhancholia S, Bhatt JC, Pant SK. 1991. Studies of some Himalayan Agarics. Acta Botanica Indica 19(1): 104-109.
- Ghosh RN, Pathak NC, Tiwari T. 1967. Studies on Indian Agaricales. Indian Phytopathol 20: 237-242.
- Holmgren PK and Keuken W. (eds) 1974. Index Herbariorum. Part I. The Herbaria of the World. Regnum Vegetabile 92: 1-397.
- Kirk PF, Cannon PF, Minter DW, Stalpers JA. 2008. Ainsworth and Bisby's Dictionary of Fungi, 10th ed. CABI Bioscience, CAB International, UK.

- Kornerup A and Wanscher JH. 1978. Methuen Handbook of Colour, 3rd ed. Eyre Methuen, London.
- Lakhanpal TN. 1993. The Himalayan Agaricales -Status of systematics. Mushroom Res 2(1): 1-10.
- Lakhanpal TN. 1995. Mushroom Flora of North West Himalayas. In: Chadha KL, Sharma SR (eds) Advances in Horticulture Vol. 13– Mushrooms, Malhotra Publishing House, New Delhi, pp 351-373.
- Manimohan PK, Thomas A, Nisha VS. 2007. Agarics on elephant dung in Kerala State, India. Mycotaxon 99: 147-157.
- Natarajan K and Raaman N. 1983. South Indian Agaricales. Bibliotheca Mycologica 89: 1-203.
- Natarajan K and Raaman N. 1984. South Indian Agaricales. A preliminary study on some dark spored species. International Books and Periodicals Supply Services, New Delhi, pp 1-204.
- Pathak NC and Ghosh RN. 1962. Fungi of Uttar Pradesh. Bulletin of the National Botanic Gardens, No. 62, National Botanic Gardens, Lucknow, India.
- Patil BD, Jadhav SW, Sathe AV. 1995. Mushroom flora of Maharashtra. In: Chadha KL, Sharma SR (eds) Advances in Hortriculture Vol. 13– Mushrooms, Malhotra Publishing House, New Delhi, pp 317-328.
- Pegler DN. 1986. Agaric flora of Sri Lanka. Kew Bulletin 12: 1-514.
- Sarbhoy AK and Daniel J. 1981. Fungi of India. CBS Publishing, New Delhi, India.
- Sathe AV and Sasangan KC. 1977. Agaricales from South West India–III. Biovigyanam 3: 119-121.
- Stamets P. 1996. Psilocybin Mushrooms of the World. Ten Speed Press, Berkeley.
- Vrinda KB, Pradeep CK, Mathew S, Abraham TK. 1999. Agaricales from Western Ghats–VI. Indian Phytopathol 52(2): 198-200.
- Wartchow F, Carvalho AS, Sousa MCA. 2010. First record of the psychotropic mushroom Copelandia cyanescens (Agaricales) from Pernambuco, Northeast Brazil. Revista Brasileira de Biociências 8: 59-60.
- Watling R and Gregory NM. 1987. British Fungus Flora-Agaric and Boleti 5. Strophariaceae and Coprinaceae. Royal Botanic Gardens, Edinburgh.